





Effect of Sodium Chloride on Ferrous Iron Oxidation and Respiratory Rate by Sulfobacillus thermosulfidooxidans

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An example of bioleaching



(Modified from Valdés, J., et. al. 2008)

(1)
$$\operatorname{CuFeS}_2 + 4 \operatorname{Fe}^{3+} \longrightarrow \operatorname{Cu}^{2+} + 5 \operatorname{Fe}^{2+} + 2 \operatorname{S}_0$$
 spontaneous
(2) $4 \operatorname{Fe}^{2+} + \operatorname{O}_2 + 4 \operatorname{H}^+ \longrightarrow 4 \operatorname{Fe}^{3+} + 2 \operatorname{H}_2\operatorname{O}$ (iron oxidizers)
(3) $2 \operatorname{S}^0 + 3 \operatorname{O}_2 + 2 \operatorname{H}_2\operatorname{O} \longrightarrow 2 \operatorname{SO}_4^{2-} + 4 \operatorname{H}^+$ (sulfur oxidizers)





 \rightarrow Chloride ions enhance leaching of copper by increasing porosity of formed sulfur layer during chalcopyrite leaching

ightarrow Chloride ions are toxic to bacteria, inhibit growth and iron oxidation activity



1. Determining the chloride effect on viability by means of the most probable number (MPN) analysis





Following 18 h exposure of 3*10⁶ cells/mL of *Sb. thermosulfidooxidans* to different NaCl levels, cells were grown in NaCl-free Mac medium.

MPN was determined with the MPN calculation program, v. 5 (Jarvis et al. 2010)

2. Determining the chloride effect bacterial growth, respiration rates and iron oxidation activity



NaCl (mM)	0	100	200	300	400	500	600	1000
Fe ²⁺ (<u>mM</u>)	50							
Yeast extract (%)	0.02							
Inoculum (%)	10							
Temperature (°C)	45							

- Determination of cell growth
- Determination of microbial respiration rates
- Determination of iron oxidation

2.1. Determining the chloride effect on total cell number of *Sb. thermosulfidooxidans* using the PicoGreen assay (*Giebner et al. 2015*)

Principle: Staining a bacterial suspension (e.g. *Sb. thermosulfidooxidans*) with PicoGreen leads to a linear response between cell numbers and relative fluorescense units (RFU)



Correlation RFU and cell number of Sb. thermosulfidooxidans

2.2. Determining the chloride effect on respiration rates of *Sb. thermosulfidooxidans* using Optode system (*Giebner et al. 2015*)

Aerobic respiration: $4Fe^{2+} + 4H^+ + O_2 \rightarrow 4Fe^{3+} + 2H_2O$



7

Results



18 h exposure to up to 200 mM NaCl cause no significant change in viable cell number

Similar exposure to 400 mM NaCl cause a decline of viable cell number by more than 3 orders of magnitude

Results

Cell growth and iron oxidation of Sb. thermosulfidooxidans in the presence of different NaCl concentrations



Results

Cell viability in comparison to *L. ferriphilum* and Fe respiration within 15 min exposure to NaCl



Fe respiration change during bacteria growth



Conclusion

- Exposure of cells to NaCl ≥ 200 mM caused an extension of lag-phase
- NaCl ≥ 300 mM strongly affected cell viability, growth and iron oxidation of Sb. thermosulfidooxidans

Outlook

- Effect of NaCI on sporulation of Sb. thermosulfidooxidans
- Effect of NaCl on Sb. thermosulfidooxidans in dependence of other substrates (sulfur, pyrite)
- Biofilm formation and Bioleaching of metal sulfides by *Sb. thermosulfidooxidans* in the presence of tolerated conc. of NaCl
- Metabolomics response of NaCl stress in Sb. thermosulfidooxidans