


# Microbial abundance and transport in glacial near-surface meltwater

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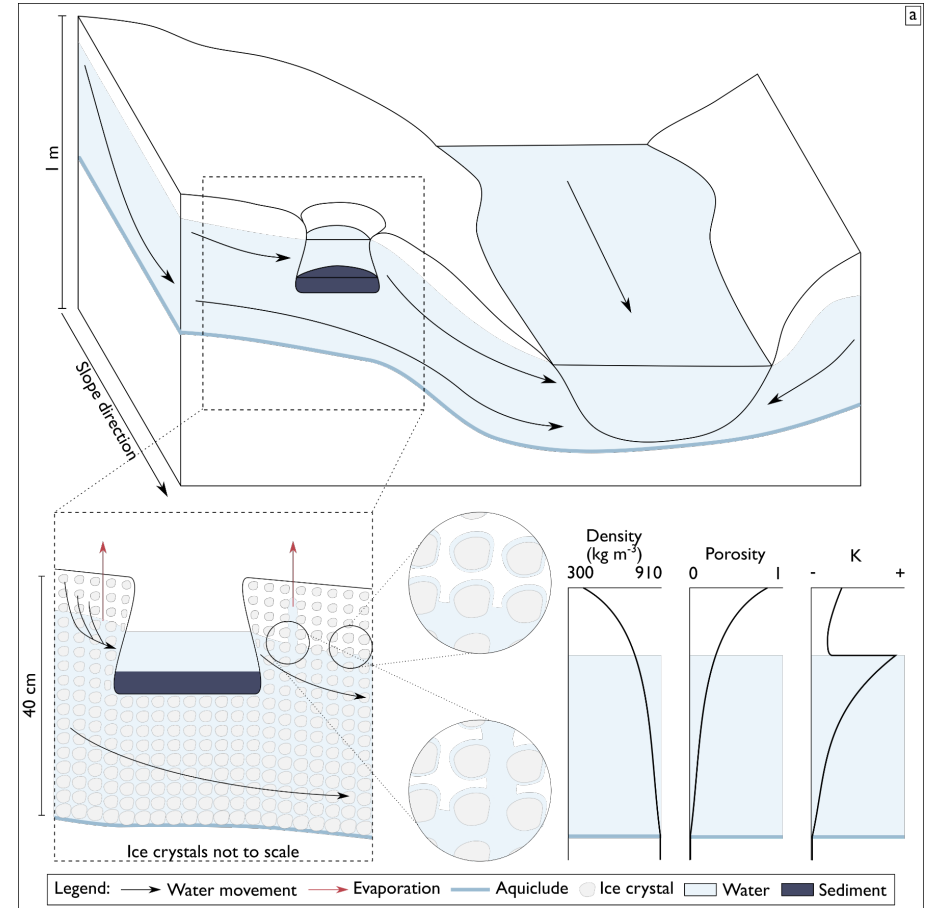
# Acknowledgements

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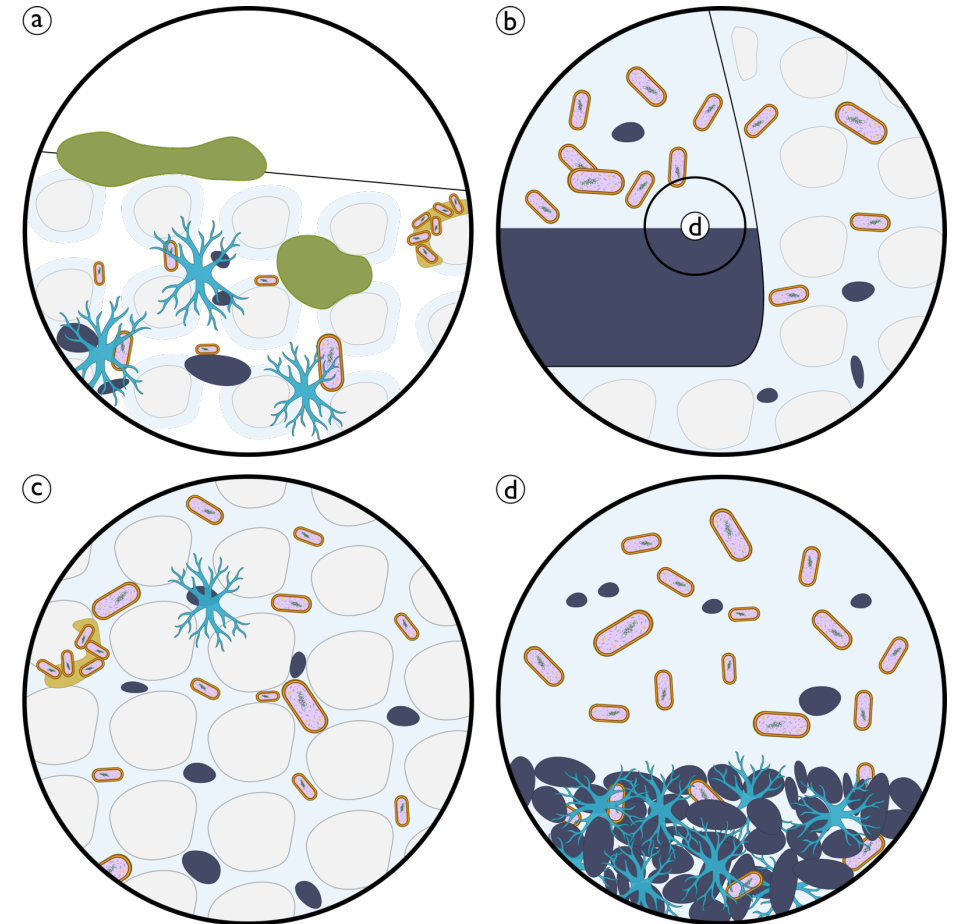
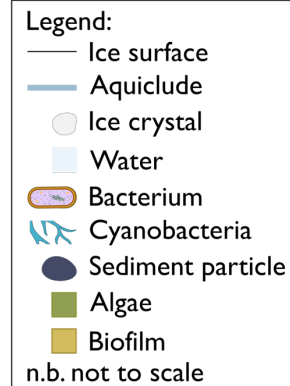
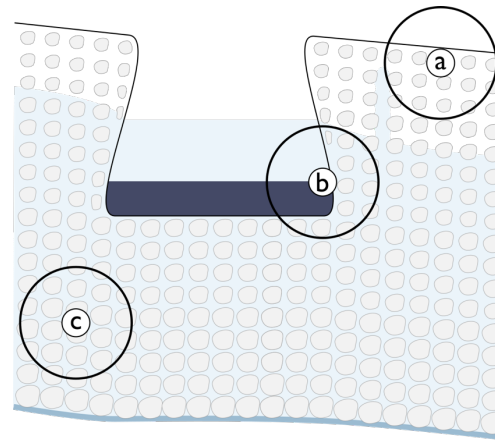
# Background

- Near-surface ice melts internally due to SWR penetration
- Preferential melt along ice crystal boundaries
- SWR penetration is depth limited
- Reduction of bulk density and enhancement of primary porosity of ice < 0.4 m deep
- Meltwater fills pore space in upper 0.4 m
  - A non-weathered ice aquiclude prevents further downward movement of water
- Perched aquifer ➤ “weathering crust”
- Pathway for meltwater into supraglacial stream network



# Background

- Light + water makes the WC a “photic zone”
- Multitude of microbial habitats
  - Cryoconite most well known
  - Also planktonic and ice bound
- Active microbial communities with auto- and heterotrophs
  - Eukaryotes, prokaryotes and viruses
- $\leq 10^5$  cells mL<sup>-1</sup>; c.  $10^{25-29}$  globally
  - Similar to pedosphere ( $10^{29}$ )



# Rationale and aim

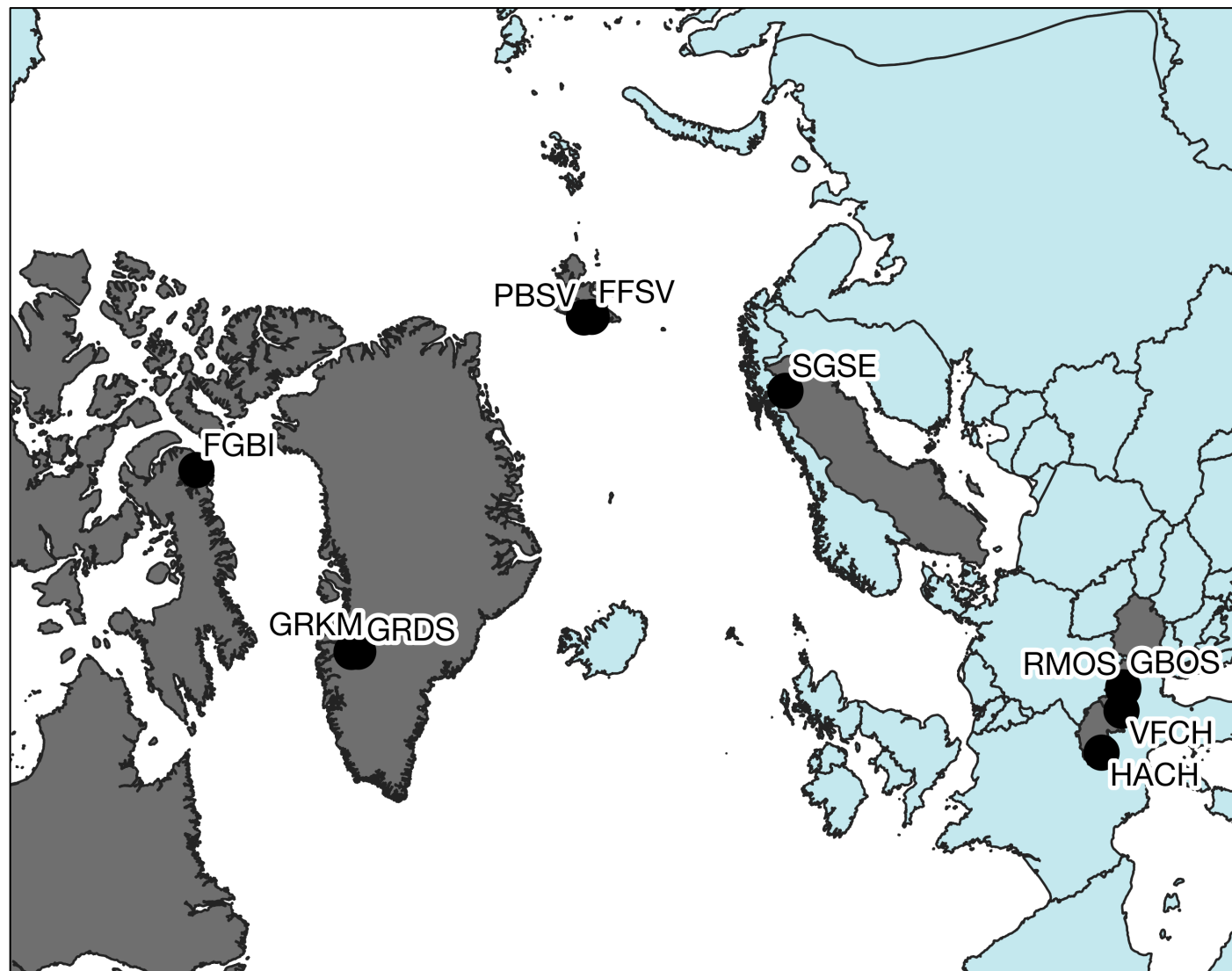
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- Current glacial microbial enumeration estimates are based on a limited number of samples when compared with other ecosystems
  - Limited understanding of controls on planktonic cell concentrations
  - Unconsidered reservoir of carbon
  - Links with bioalbedo of ice surfaces
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- **AIM: enumerate microbes in weathering crust waters across the Northern hemisphere and explore potential hydrological controls on community numbers**

# Methods: Field sites

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- 10 glaciers in 6 countries including a range of geographic locations, latitudes and climatic settings
- 2014 – 2016 ablation seasons



# Methods: Data collection

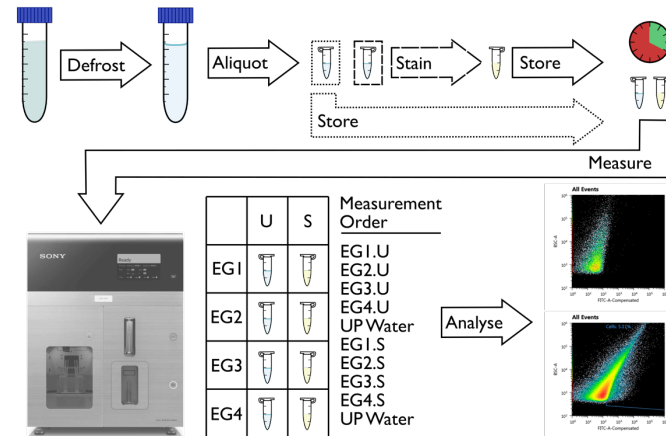
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- WC hydraulic conductivity (k) recorded via bail-recharge of auger holes (see Stevens *et al.*, 2018 Hydro. Proc.)
- Stream discharge via salt dilution at PBSV, FFSV, FGBI, SGSE, HACH and VFCH
- Electrical conductivity and temperature of WC water at PBSV, HACH and VFCH
- Paired collection of water samples for microbial enumeration using sterile 15 mL centrifuge tubes and pre-contaminated equipment
- Samples fixed with 2 % w/v paraformaldehyde (VFCH) or glutaraldehyde (others), kept in the dark and frozen within 8 hours of collection
  - Stored at -80 °C within three weeks of collection until analysis

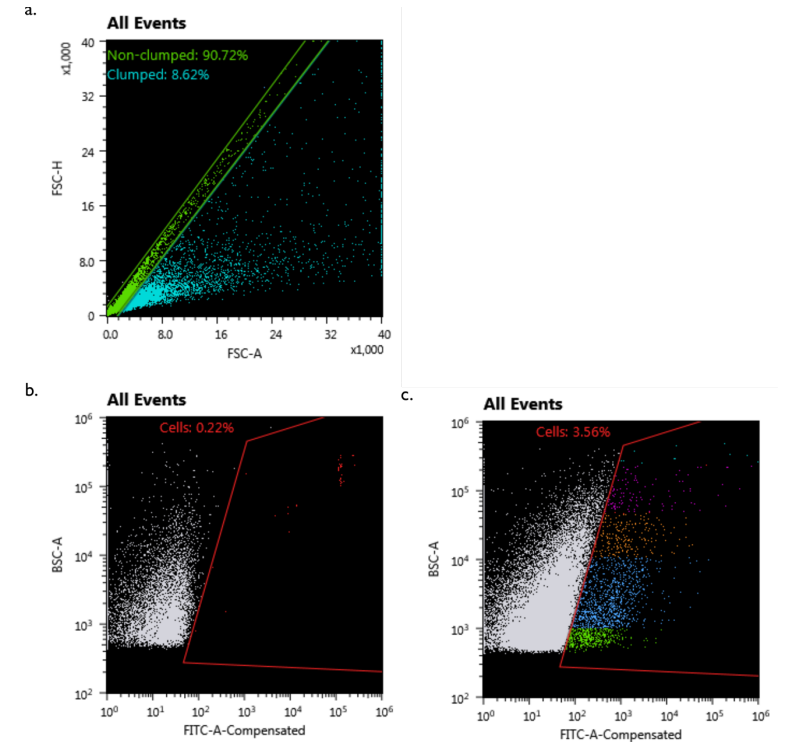


# Methods: Microbial enumeration

- Flow Cytometry (FCM) using a Sony SH800-EC
- Samples defrosted overnight
- Stained using SYBR Gold DNA stain
- Comparison of stained/unstained sample pairs
  - Quality control to ensure that cells and sediment are not “clumped”
  - Stained material = cells
- “Beads” of a known size used to estimate cell sizes



FCM Protocol



Gating procedure. Panel a shows the quality control process, clumped events fall outside the green area. A sample such as this would be disaggregated and reanalyzed. Panels b and c show an unstained and stained sample pair, the events (points) which shift right on the FITC axis are material stained with SYBR Gold and presumed to be microbial cells.

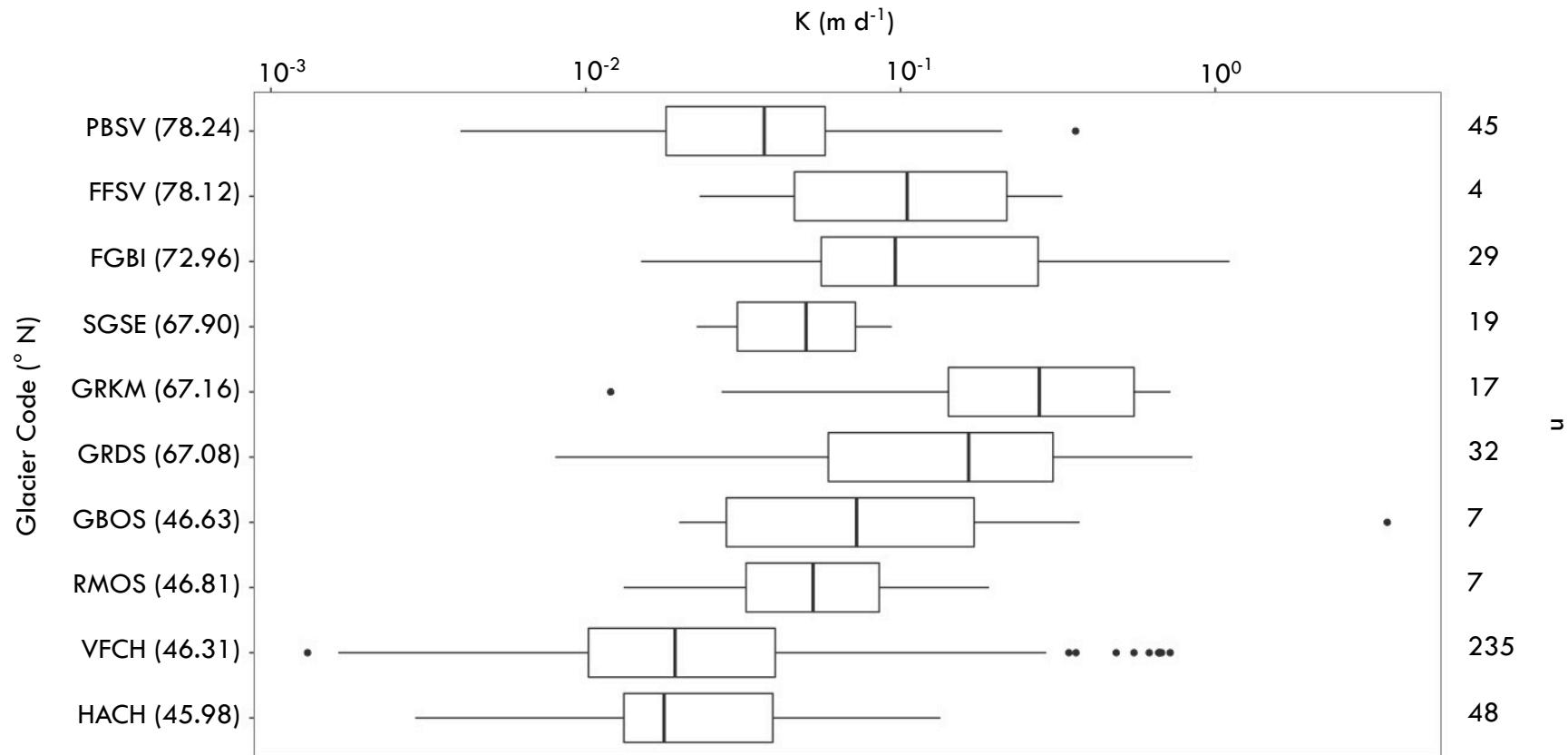


# Methods: Carbon flux calculations

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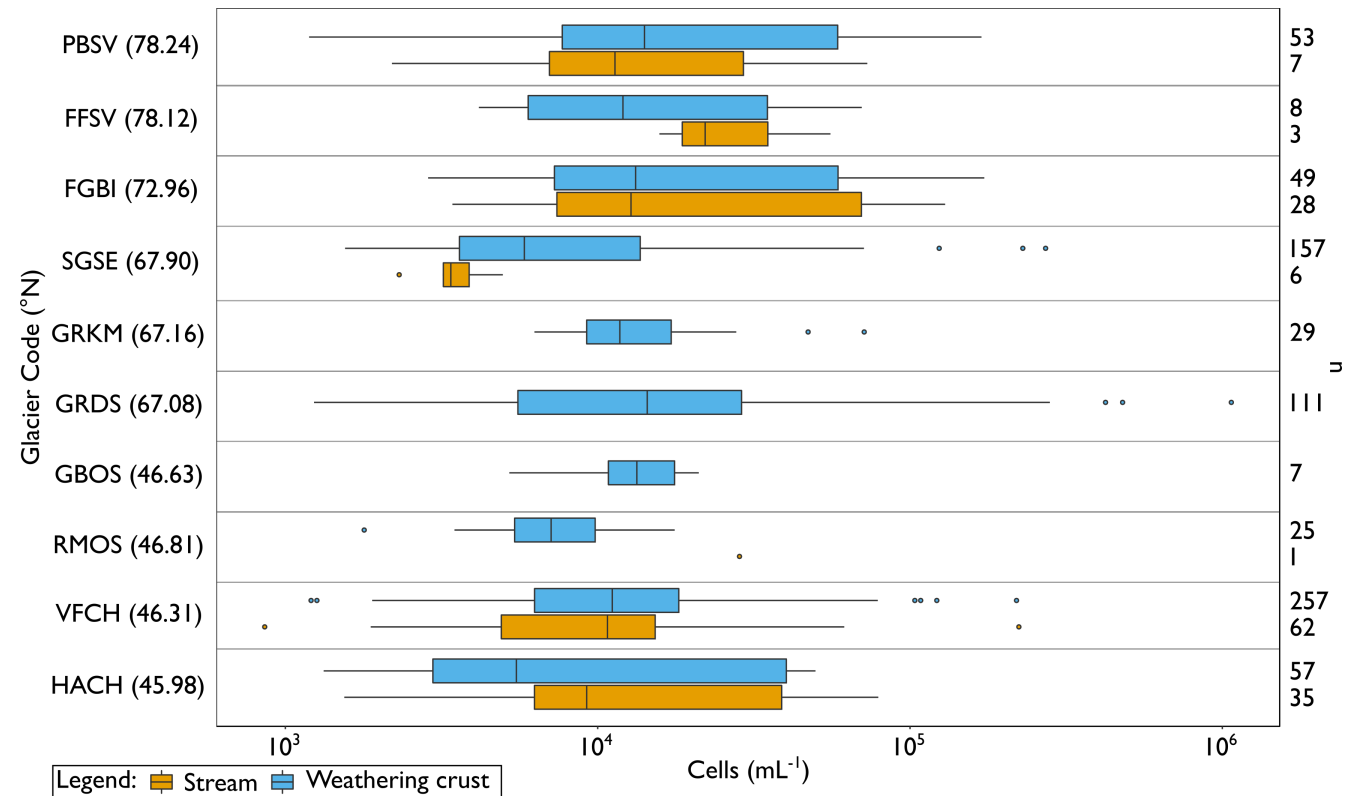
- Bacterial sizes converted to biovolume
  - Assumed cells are rod shaped: cylindrical with semi-hemispheric ends
  - Mid-point of size class used as length
  - Standard geometries (relative to length) to calculate diameter and volume
- Biovolume  $\rightarrow$  C using a conversion factor
  - $5.6 \times 10^{-13}$  g C  $\mu\text{m}^{-3}$  (Bratbak, 1985)
- Runoff estimates under RCP 4.5 to 2099 from Bliss *et al.* (2014)
- Underlying assumptions that runoff:
  - Originates on the surface
  - Transports all entrained microbes to the pro-glacial fluvial/marine system

# Results: Hydraulic conductivity



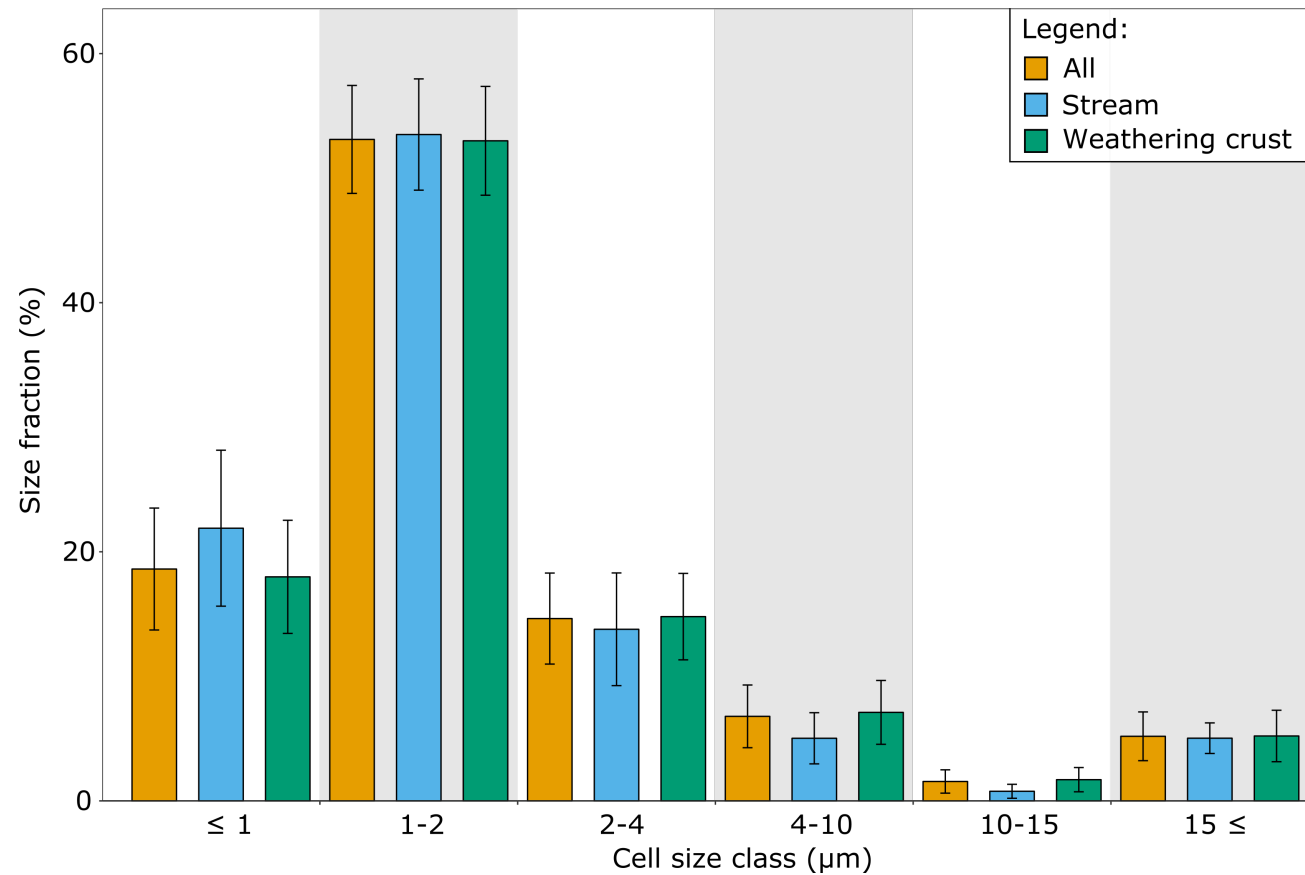
Median  $K = 2.8 \times 10^{-2} \text{ m d}^{-1}$  ( $n = 443$ )

# Results: Microbial enumeration

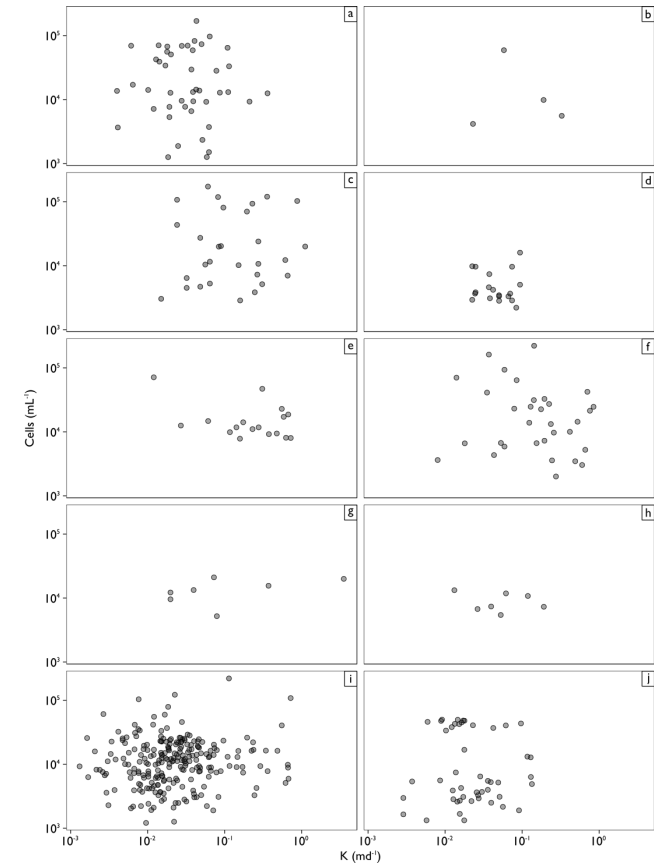
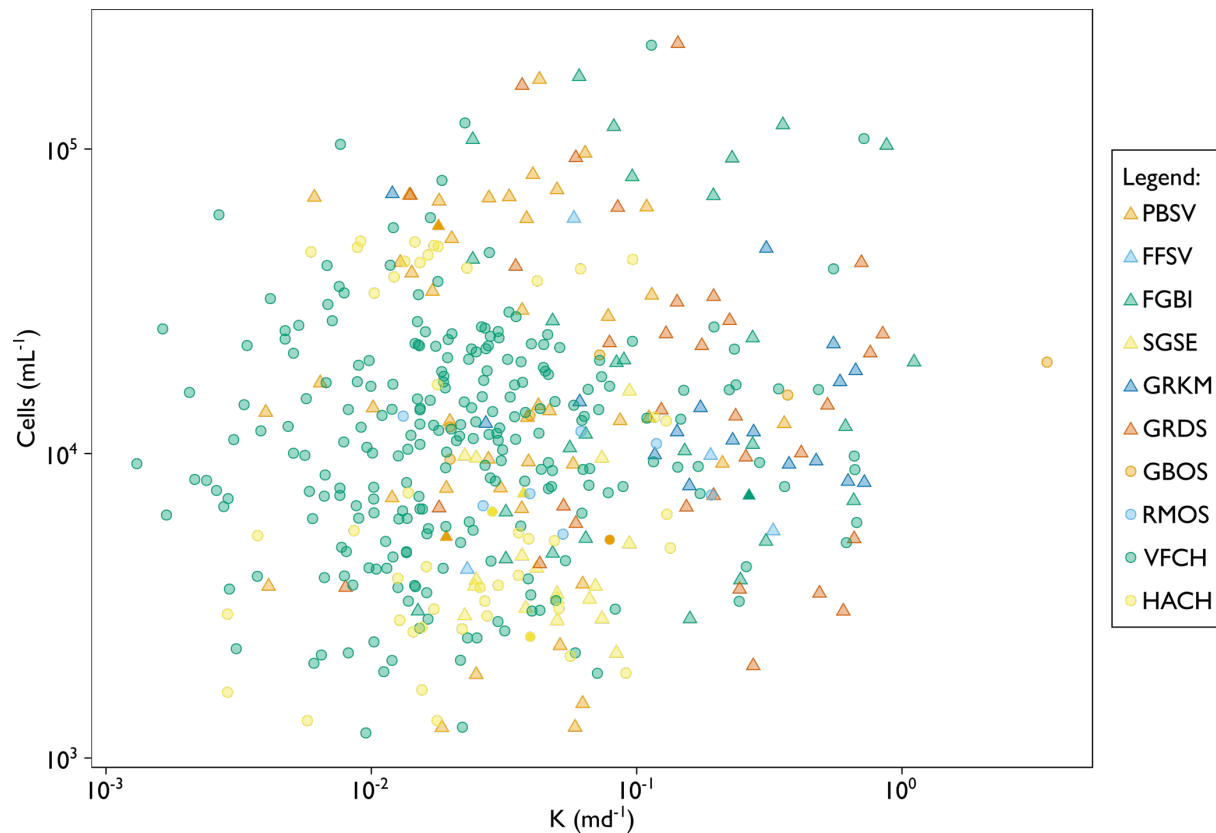


Mean microbial abundance =  $2.2 \times 10^4$  cells mL<sup>-1</sup> (n = 895)

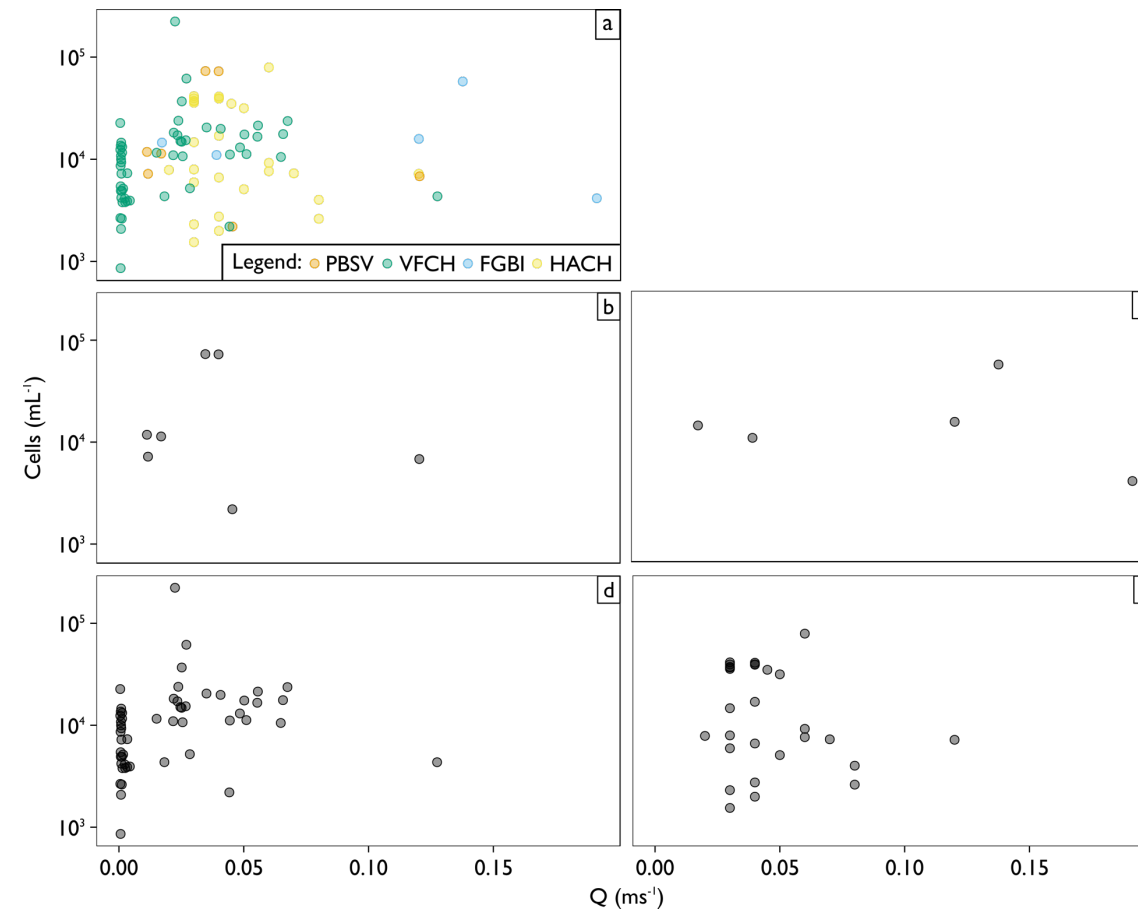
# Results: Microbe size



# Results: hydrology as a control on cells – weathering crust



# Results: hydrology as a control on cells – streams



# Results: Carbon export from supraglacial meltwater

Region	2003 – 2022 (kg a <sup>-1</sup> )	2041 – 2060 (kg a <sup>-1</sup> )	2080 – 2099 (kg a <sup>-1</sup> )	2003 – 2099 (kg a <sup>-1</sup> )
Svalbard	$4.8 \times 10^7$	$6.1 \times 10^7$	$4.3 \times 10^7$	$5.2 \times 10^7$
Arctic Canada (N)	$9.8 \times 10^7$	$1.3 \times 10^8$	$1.3 \times 10^8$	$1.2 \times 10^8$
Greenland	$1.6 \times 10^8$	$1.7 \times 10^8$	$1.6 \times 10^8$	$1.7 \times 10^8$
Scandinavia	$4.1 \times 10^6$	$3.0 \times 10^6$	$1.5 \times 10^6$	$3.0 \times 10^6$
Central EU	$2.7 \times 10^6$	$1.5 \times 10^6$	$8.9 \times 10^5$	$1.8 \times 10^6$
Total	$3.7 \times 10^8$	$3.7 \times 10^8$	$3.7 \times 10^8$	$3.4 \times 10^8$
<b>Global Total</b>	<b><math>1.2 \times 10^9</math></b>	<b><math>1.2 \times 10^9</math></b>	<b><math>9.9 \times 10^8</math></b>	<b><math>1.1 \times 10^9</math></b>

Regions are split as follows: Svalbard: FFSV, PBSV; Arctic Canada: FGBl; Scandinavia: SGSE; Greenland: GRDS, GRKM; Central EU: GBOS, HACH, RMOS, VFCH.

Regions sampled represent  $\approx 30\%$  of modelled global glacier runoff over the next century, which is upscaled to provide the global total. Reduction in C export from the 2003-2022 period to the 2080-2099 period is solely a function of discharge reduction as glaciers retreat.

# Cell numbers and size

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- Lower-end estimate of supraglacial microbes as only those in meltwater sampled
- Mean concentrations align with those in other studies
  - Fall within the range found in terrestrial groundwater aquifers ( $10^2 - 10^8$  cells mL<sup>-1</sup>)
  - Similar range to snow ( $< 10^5$  cells mL<sup>-1</sup>)
- Significant pairwise differences between GRDS and all European Alpine glaciers (Tukey's HSD post-hoc of one-way ANOVA;  $p < 0.05$ )
  - What controls this?
- Modal cell size class of 1 – 2  $\mu\text{m}$  (slide 12)
  - Larger than size suggested by Irvine-Fynn *et al.* (2012)
  - Cells  $> 15 \mu\text{m}$  observed – could be some remaining aggregates, but could be indicative of eukaryotic cells (note that no phylogenetic data are available)



# Implications for WC hydraulics

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- No correlation between K or Q and microbial concentration (slides 13/14)
- No significant difference in microbial concentration in WC or streams (slide 10)
  - Implication that cells are advected efficiently once mobilized, not filtered by WC
  - So why does WC darken/accumulate biomass through the ablation season?
  - Possible that removal via filtering (and viral action) = addition from replication/liberation from melting ice?
- Presence of particles  $> 15 \mu\text{m}$  in meltwater implies pores of at least this diameter are present in the WC
  - Also contrasts with the idea of mechanical filtration; even large cells mobilised

# Supraglacial contribution to C flux

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- Upscaling exercise strongly controlled by runoff estimates
  - Designed to highlight potential role of glacier surfaces in C cycles
  - Uncertainties in runoff estimates incorporated
  - No consideration of surface cover changes – e.g. increasing debris cover with ablation
  - Excludes cells bound to ice which may be released during WC degradation events
- Implied liberation of  $10^9$  kg C  $\text{a}^{-1}$  as POC over next century (slide 15)
  - $\approx 0.08$  % of aquatic biosphere
- 44.6% of water and therefore C flux is from mass loss rather than annual cycles; implies the liberation of “locked-up” C from within ice

# Summary

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- Median WC K of  $2.8 \times 10^{-2} \text{ m d}^{-1}$  (similar to a sandstone)
- Mean microbial concentration of  $2.2 \times 10^4 \text{ cells mL}^{-1}$  in surface meltwater
  - Highest in Greenland, lowest in European Alps
- No evidence for mechanical filtration by the WC
  - Particles  $> 15 \text{ }\mu\text{m}$  mobilised
- $10^9 \text{ kg C a}^{-1}$  released downstream from glacier surfaces to fluvial/marine environments
- However, lots of questions unanswered...

# Future Work

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- Explore other potential controls on microbe concentration:
  - Nutrient input and bioavailability?
  - Cell replication and viral control rates?
  - “Age” of the weathering crust?
- Darkening of the surface throughout the melt season
  - Role of ice-bound microbiota?
  - Tracer studies to determine flow paths: throughflow or microbe “input” = “output”?

