# Multi-disciplinary approach for assessing the impact of a flood event in a shallow karst cavity (Pindal Cave, Spain)

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#### **1. Introduction**

Pindal Cave (Asturias, Spain) and its Paleolithic art have been part of the UNESCO World Heritage List since 2008. Since 2017, a multidisciplinary team have been carrying out deciphering the research focused on relationships between environmental conditions and microbial activity to the design



Fig. 1. Paleolithic art and Pindal Cave location. of conservation strategies. On October 23, 2019, the cave was flooded. After the flood, a specific study was performed to evaluate the environmental changes caused in the underground ecosystem. This study revealed a massive entry of material from a cattle farm located vertically in the cavity. Thereafter, in 2021, the livestock activity in the vicinity of the cave was stopped.

### 2. Why did the flood occur?

The Pindal karstic system develops in a calcareous massif (Carboniferous) modeled in the form of an erosional marine terrace (rasa) by coastal morphogenetic processes. This marine terrace level is located at an elevation of 30-68 meters above current sea level and constitutes the preferred catchment area for runoff water from another higher level (140-170 m) developed on quartzite layers with very low permeability (Ordovician). The cave is the main endokarstic feature of the system. On the surface of the 30-68 m rasa there are numerous exokarstic structures of sinkhole and polje type. On one of the sinkholes, located almost vertically to the cave, a cattle farm was installed in 1995 (Fig. 2).

Between October 19 and 23, 2019, an extreme episode of rainfall occurred in the area with a cumulative total of 209 l/m<sup>2</sup>. This event caused a strong accumulation of water in the sinkhole that finally collapsed, flooding the cave for several days (Fig. 3).

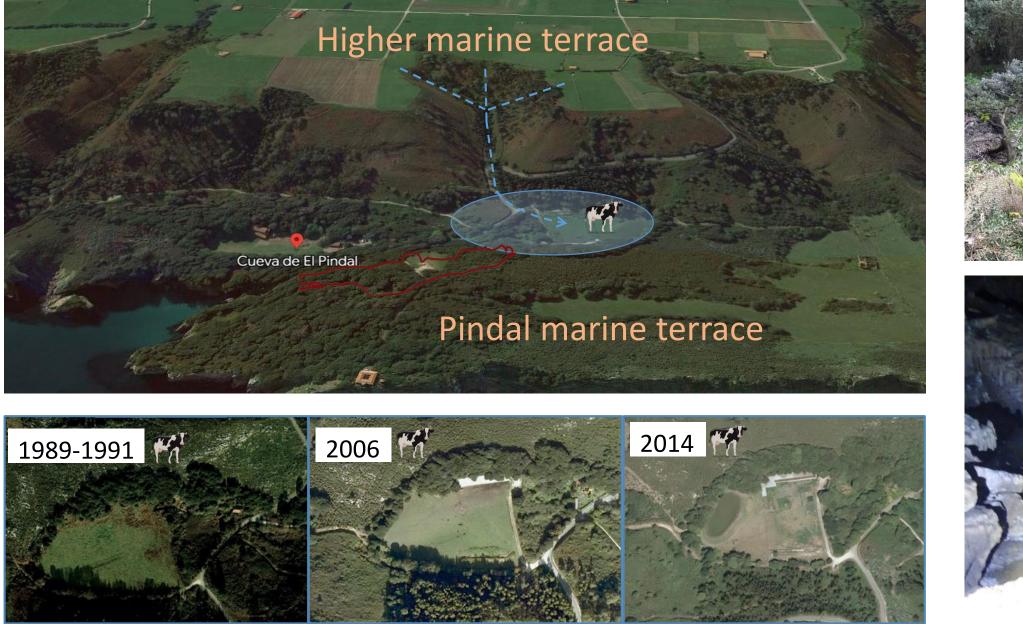


Fig. 2. Aerial view of the rasas (top) and evolution of the farm (bottom). Blue indicate the discharge area.



Fig. 3. Collapsed sinkhole (top) <sup>[1]</sup> and flooded cave (bottom).

#### References

<sup>[1]</sup> Jennings, J.N., 1985, Karst Geomorphology: Oxford, Basil Blackwell, 293 p.

<sup>[2]</sup> Martin-Pozas, T., Sánchez-Moral, S., Fernández-Cortes, A., et al. (2020). Biologically mediated release of endogenous N<sub>2</sub>O and NO<sub>2</sub> gases in a hydrothermal, hypoxic subterranean environment. Science of the Total *Environment*, 747, 141218. https://doi.org/10.1016/j.scitotenv.2020.141218.

<sup>[3]</sup> Martin-Pozas, T., Cuezva, S., Fernandez-Cortes, A., et al. (2022). Role of subterranean microbiota in the carbon cycle and greenhouse gas dynamics. *Science of the Total Environment*, 831, 154931. https://doi.org/10.1016/j. scitotenv.2022.154921.

## 3. Methods

One week after the flood, the sediments were sampled at three points for their biogeochemical subsequent and microbiological analysis <sup>[2,3]</sup>. The results were compared with those obtained before and one year after the flooding. For microbiological analysis, DNA was extracted from moonmilk, cave and exogenous sediments. Then, 16S rRNA gene amplicon sequencing was performed on Illumina, and reads were analyzed using QIIME 2 and the SILVA 132 database <sup>[3]</sup>.

4. Geochemical results **5.** Microbiological results The cave sediments and exogenous sediments of sites 7 and 9 showed high One week after the flooding the exogenous sediments (In Fig. 8: 9-1 in red) showed high values of organic matter, nitrogen phosphorus and metals. These values relative abundances of Thauera, Corynebacteriaceae, Bacillota, Bacteroidota and were higher than those of the innermost sites (17) that were not directly methanogenic archaea characteristic of the gastrointestinal tract and contaminated affected by the collapse of the sinkhole and the entry of water contaminated environments. After Before Flooding with slurry. Subgroup 2-Phylum (Bacteria) Subgroup 9 • Acidobacteriota P (mg/kg) **Organic matter (%)** Total N (%) Actinomarinales -Actinomycetota Corynebacteriaceae 0<mark>.3</mark>4 Racteroidota Pseudonocardiaceae 23<sup>24</sup> Bacteroidetes -0.30 Angerolinege 6 7 66 0.09 Dehalococcoidia 0.04 21 6 5 JG30-KF-CM66 (Alpha) 'Delta) 'Gamma) Firmicutes Gemmatimonadaceae Nitrospira 🛛 Before 💻 Flooding 🖿 After 📄 Before 💻 Flooding 💻 After Before 📕 Flooding 🔲 After Gemmataceae Pirellulaceae Pb (mg/kg) Cu(mg/kg) Zn (mg/kg) Beijerinckiaceae · Pedomicrobium 65<sub>c</sub>, Paracoccus 33 NB1-j \_28 1213 Thauera **Relative** abundance Nitrosomonadaceaemost abundant reads Nitrosococcaceae PLTA13 ite 9. Before: before Methylomirabilaceae oding, Flooding: one Rokubacteriales the flooding, Before EFlooding After Before Flooding After Eflooding Flooding Flooding After after Nitrosopumilaceae Nitrososphaeraceae one year after the Fig. 6. Results of the geochemical analysis. Before: before the flooding, Methanomassiliicoccales flooding. 9-1 corresponds Methanomethylophilaceae -Flooding: one week after the flooding, After: one year after the flooding. 100 to layer 1 and 9-2 to layer 2 Methanosarciniales-Red: sediments contaminated with slurry (sites 9 and 7). of Fig. 4. 9-2 9-1 9-2



Maroon: uncontaminated sediments (site 17).

### 6. Conclusions

- of microbial populations in the cave.
- the intestinal tract of mammals.



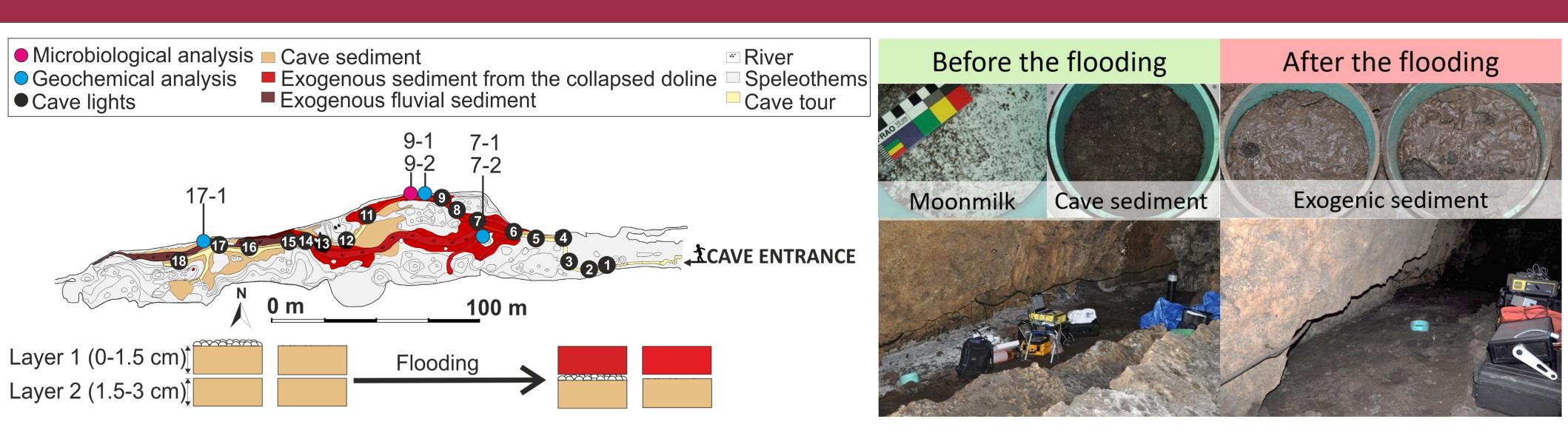
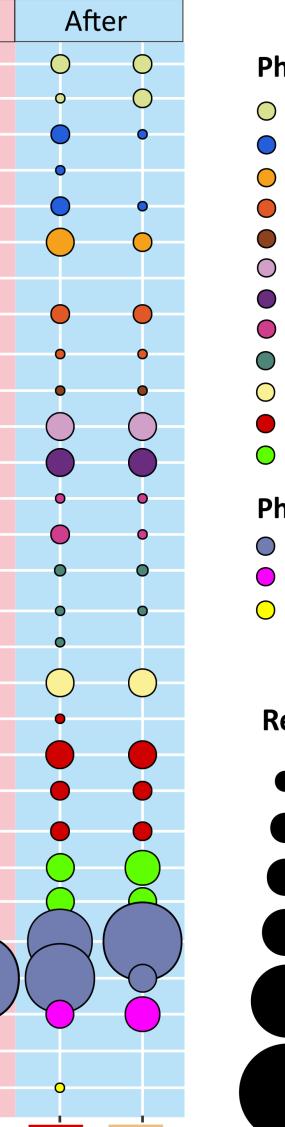


Fig. 4. Flood affected areas and sampling strategy.

• The entry of contaminated water was located at site 13 and affected the areas inside the cavity located between site 6 and site 16 (red zone in Fig. 4). • The influence of livestock activity in the cave is not recent, it has taken place over a prolonged time and results in high organic matter content that favors the development

The flooding modified the microbial communities of the affected areas, introducing genetic material from anaerobic bacteria and methanogenic archaea characteristic of

Fig. 5. Microbiological sampling area (site 9).



$\bigcirc$	Bacteroidota		
$\bigcirc$	Chloroflexota		
	Bacillota		
$\bigcirc$	Gemmatimonadota		
	Nitrospirota		
$\bigcirc$			nycetota
$\bigcirc$	Pseudomonadota (A		
$\bigcirc$	Pseudomonadota (I		
	Pseudomonadota (C		
$\bigcirc$	Me	thylo	mirabilota
Phylum (Archaea)			
$\circ$	Thermoproteota		
$\bigcirc$	Thermoplasmatota		
$\bigcirc$	Halobacteriota		
$\bigcirc$	1101	CNUC	
$\mathbf{D}_{\alpha\alpha}$ de $(0/)$			
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