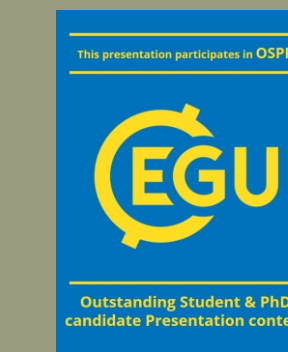




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Testing the use of Sodium Polytungstate for an efficient extraction of microplastic particles from river sediments

Francesca Uguagliati¹, Massimiliano Zattin¹ and Massimiliano Ghinassi¹
¹Department of Geosciences, University of Padova, Italy



EGU23-6498
francesca.uguagliati@phd.unipd.it
f_uguagliati

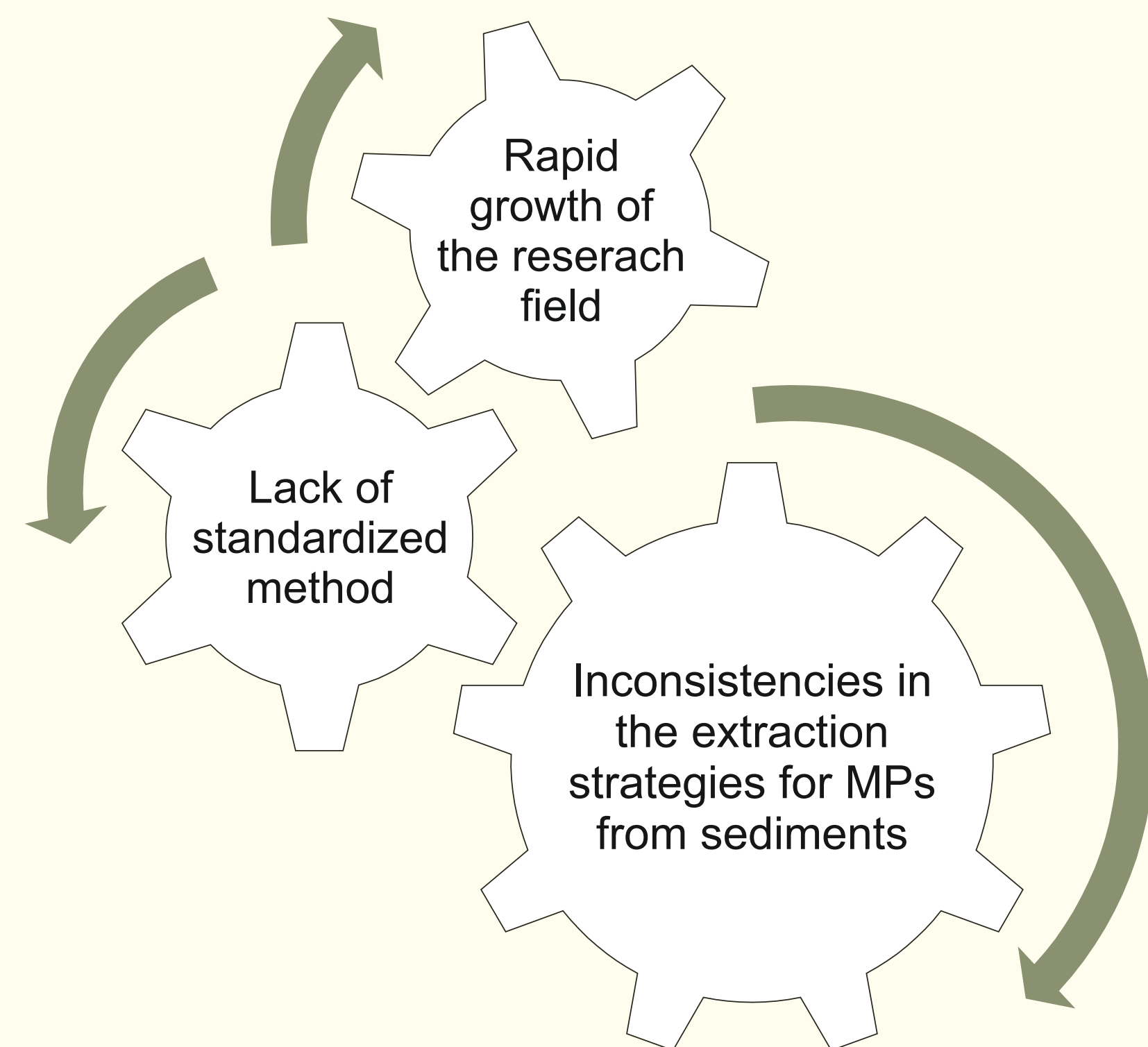
Introduction

Rivers:

- Primary pathways of MPs to the sea^{1,2,3},
- Reservoirs of MPs stored in sediments^{4,5}.

Kowing microplastics content in river sediments is critical.

The optimization of a methodology for the separation and classification of microplastics from sediments is crucial.



Several studies highlight that density separation using a high-density concentrated saline solution is one of the most reliable and efficient separation methods^{6,7,8,9}.

Aim of the Study

Testing the use of Sodium Polytungstate as a density separation agent.

What is the recovery rate of this method?

References

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Methodology

1. Creation of artificial sediments

20-25 grams of sediment plus 50 MPs per sample for a total of 52 samples

Plastic-free sediments
(Pleistocene alluvial deposits)



Used particle size ranges

- Mixture A: 63 - 250 μm
- Mixture B: lower than 63 μm
- Mixture C: 50% A + 50% B

Selected & added microplastic particles



PET films
($\rho = 1.38 \text{ g} \cdot \text{cm}^{-3}$)



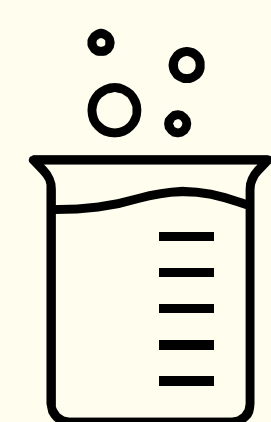
PET fibres
($\rho = 1.38 \text{ g} \cdot \text{cm}^{-3}$)



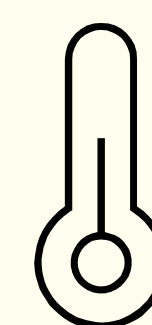
PVC fragments
($\rho = 1.4 \text{ g} \cdot \text{cm}^{-3}$)

2. Organic matter removal

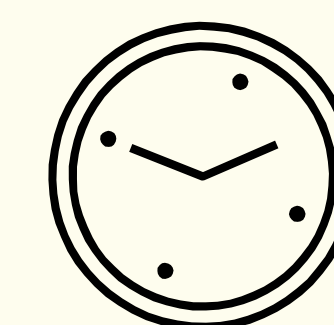
Wet Peroxide Oxidation^{10,11}



$\text{H}_2\text{O}_2 + \text{Fe}^{2+}$



50 - 70 $^{\circ}\text{C}$



< 4 h

3. Density separation

Sodium Polytungstate (ρ from $3.1 \text{ g} \cdot \text{cm}^{-3}$ to $1.6 \text{ g} \cdot \text{cm}^{-3}$)

- Indermediate density between sediments and plastic^{12,13},
- Non-toxic^{12,13},
- Easily recoverable and reusable^{12,13},
- Successfully used for the separation of minerals and rocks¹³.

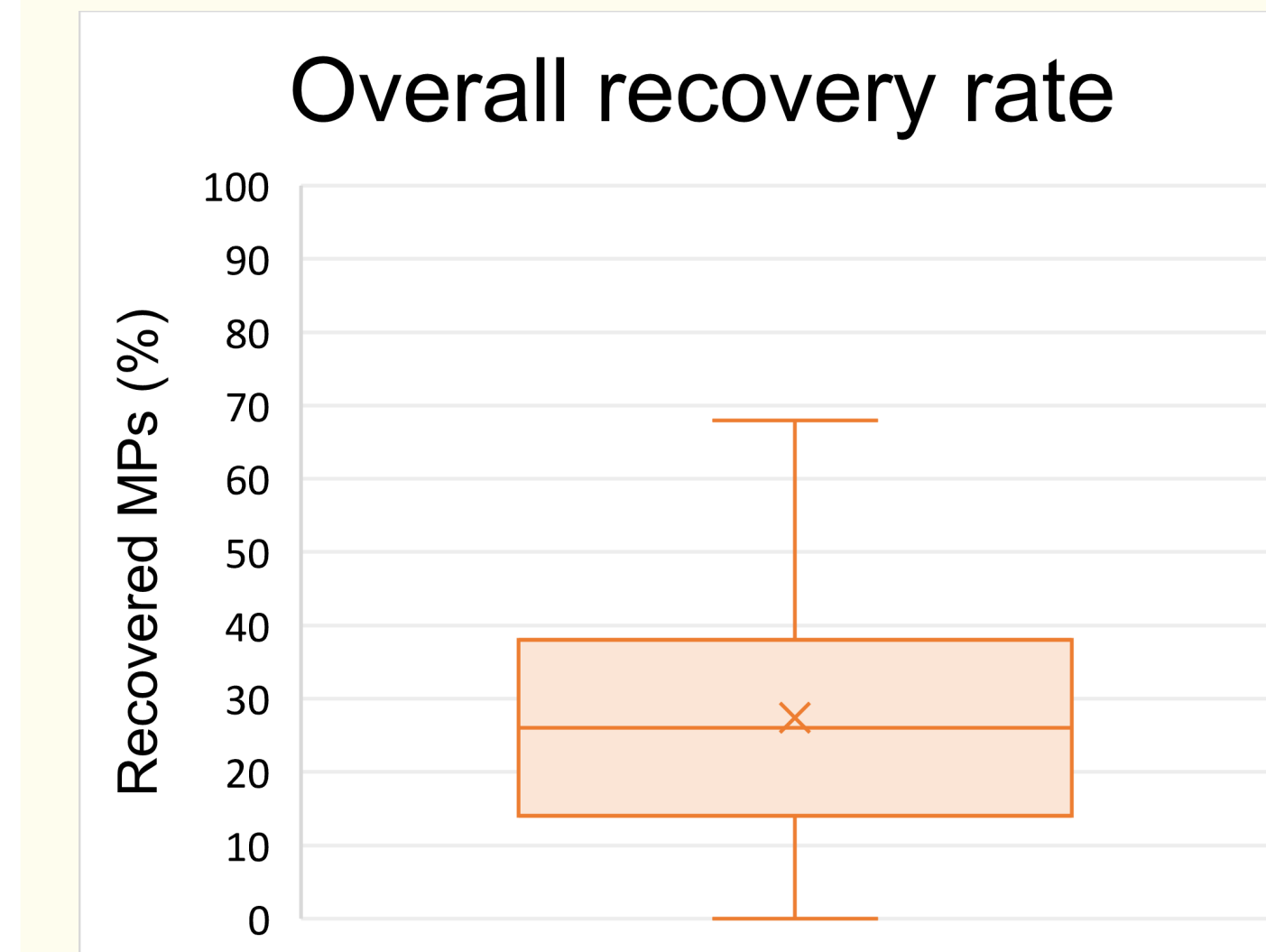
The procedure:

- Addition of the liquid to the samples, centrifugation and settling,
- Collection of the suspended particulate matter,
- Transfer of the suspension to cellulose filters,
- Microplastics counting under a stereomicroscope.



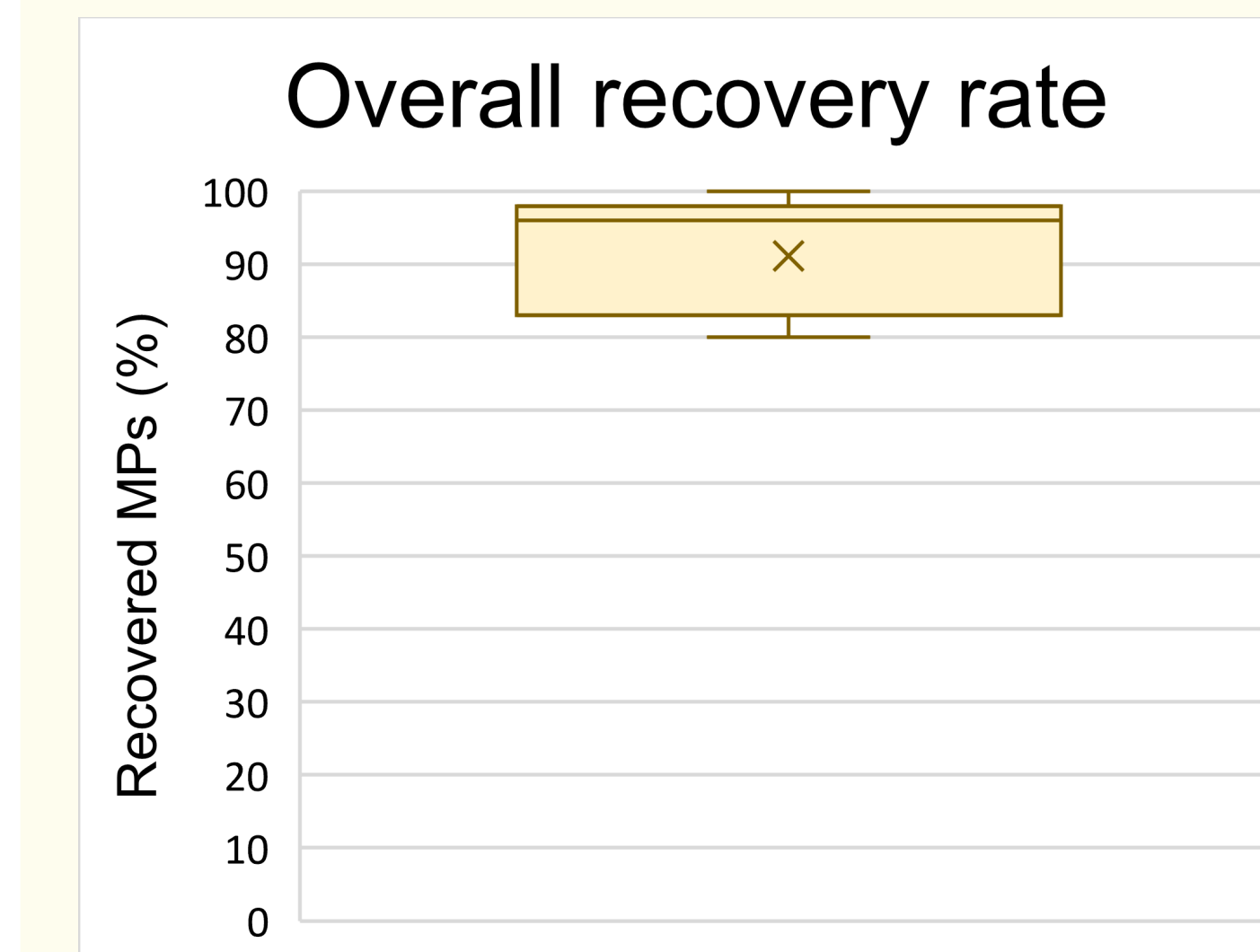
Preliminary Results

Procedure 1



- Test tube capability: 50 ml,
- 25 g of sediments,
- 50 microplastic particles.

Procedure 2



- Test tube capability: 500 ml,
- 20 g of sediments,
- 50 microplastic particles.

Conclusions

Procedure 1

Low recovery rate due to:

- MPs trapped in sediments,
 - Test tube capability (50 ml) vs the amount of sediments (25 g).
- MPs dispersion during different steps.
- Too strong organic matter removal,
 - Destruction of the several layers of wich glitters are made of.

Procedure 2

High recovery rate due to:

- Use of the same container for all the steps,
- Larger containers: 500 ml,
- Lower amount of sediments (20 g).